

Best Practice & Research Clinical Obstetrics and Gynaecology

Contents lists available at [ScienceDirect](www.sciencedirect.com/science/journal/15216934)

BEST

journal homepage: www.elsevier.com/locate/bpobgyn

10

Immunotherapy of gynecological cancers

Emad Matanes, MD, Walter H. Gotlieb, MD, PhD *

Division of Gynecologic Oncology, Segal Cancer Center, Lady Davis Institute of Research, Jewish General Hospital, McGill University, Montreal, Canada

Keywords: Immunotherapy Checkpoint inhibitors Vaccine Ovarian cancer Cervix cancer Endometrial cancer

ABSTRACT

Oncology treatments have evolved from intuitive, via empiric, to the present precision medicine, with the integration of molecular targeted therapies in our treatment arsenal. The use of the patients' powerful immune system has long been contemplated and recently led to the integration of immunotherapy to overturn the well-documented inhibitory effects of the tumor on the immune system and restore it to a state of activity against the cancer. Recent favorable results have shown the value and effectiveness of immunotherapy against gynecological cancers. In particular, the checkpoint inhibitors, targeting the programmed death-1 (PD-1) pathway, have shown durable clinical responses with manageable toxicity. Several phase II and III clinical trials testing the association of different regimen of chemotherapy and immunotherapy are ongoing in gynecological cancers, and important results are expected. In this chapter, we outline the main principles of immunotherapy for gynecological cancers and summarize the current strategies used in clinical trials.

© 2019 Elsevier Ltd. All rights reserved.

Index

Innate immunity: Cells and mechanisms that provide the first line of defense in a nonspecific manner. Innate immune responses are rapid and independent of antigen [\[1\]](#page-10-0).

E-mail address: walter.gotlieb@mcgill.ca (W.H. Gotlieb).

<https://doi.org/10.1016/j.bpobgyn.2019.03.005>

^{*} Corresponding author. Division of Gynecologic Oncology, McGill University, SMBD Jewish General Hospital, 3755 Chemin de la Cote-Ste-Catherine, Montreal, QC, H3T 1E2, Canada.

^{1521-6934/}© 2019 Elsevier Ltd. All rights reserved.

Adaptive immunity: Cells and mechanisms that provide protection that is mediated by B and T lymphocytes following exposure to specific antigen and characterized by immunological memory [\[2\].](#page-10-0)

Humoral immune response: Transferred by serum. Refer to antibody responses; antibodies are antigen-reactive, soluble, bifunctional molecules composed of specific antigen-binding sites associated with a constant region that directs the biologic activities of the antibody molecule, such as binding to effector cells or complement activation [\[3\].](#page-10-0)

Cellular immune response: Transferred by cells. Generally, refer to cytotoxic responses mediated directly by activated immune cells, rather than by the production of antibodies T lymphocytes: act as helper cells in the generation of humoral and cellular immune responses and by acting as effector cells in cellular responses [\[4\].](#page-10-0)

T helper/inducer cells: Express the CD4 cell surface marker, play a main rule in the production of cytokines, and can provide help to B lymphocytes, resulting in antibody production [\[5\]](#page-10-0).

T suppressor/cytotoxic cells: express the CD8 marker and can directly kill target cells [\[6\]](#page-10-0).

Major Histocompatibility Complex (MHC): Both CD4 and CD8 T cells respond to antigen only when it is presented in the context of major histocompatibility complex (MHC) molecules on antigenpresenting cells, target cells, or both [\[7\]](#page-10-0).

B lymphocytes: Cells that produce and secrete antibodies, which are antigen-binding molecules. They also can serve as efficient antigen-presenting cells for T lymphocytes [\[8\].](#page-10-0)

Macrophages and dendritic cells (DCs): Play a key role in the generation of adaptive, lymphocytemediated immune responses by acting as antigen-presenting cells [\[9\].](#page-10-0)

Natural killer (NK) cells: Have large granular lymphocytic morphology, do not express the CD3 T cell receptor complex, and do not respond to specific antigens [\[10\].](#page-10-0)

Cytokines: Soluble mediator molecules that induce, enhance, or affect immune responses [\[11\]](#page-10-0).

Introduction

Cancers are more prevalent in populations who are immunosuppressed (e.g., patients with HIV and organ transplant recipients), suggesting that impaired immunity may contribute to the development of cancer [\[12\].](#page-10-0) Considering the immense potential of the immune system to fight "foreign," many efforts for more than 35 years have attempted to develop strategies for actively stimulating immunological rejection of tumors. In 2017, the Food and Drug Administration (FDA) approved the first two chimeric antigen receptor-T cell therapies for the treatment of patients with refractory or relapsed B-cell leukemia [\[13\].](#page-10-0)

This chapter will outline the main principles of immunotherapy for gynecological cancers and summarize the current strategies used in clinical trials.

Therapeutic strategies

Immunotherapy is utilized to overturn the well-documented inhibitory effect of the tumor on the immune system and restore it to a state of activity. The tumor is able to elude immune response through multiple pathways, including the activation of immune checkpoints that inhibit the immune system's ability to target the tumor [\[14,15\]](#page-10-0). To activate tumor-directed immune responses, recent immune therapies have utilized several approaches:

1 **Adoptive cell transfer (ACT):** Autologous T cells, meaning the T cells from the patients, undergo either ex-vivo extraction from resected tumor tissue (tumor-infiltrating lymphocytes (TILs)) or from peripheral blood from the patient, and are then expanded in vitro, and infused back into the patient with the purpose that these T cells will attack the cancer cells. In addition, to improve specificity against the cancer, genetically modified T cells, designed to recognize specific tumor-associated antigens, may be included into adoptive cell transfer. These include engineered T-cell receptors that recognize tumor-specific peptides or chimeric antigen receptors (CAR) that allow T cells to recognize tumor surface antigens using an antibody-like recognition module. This technology has provided the best results, to date, in tumor immunotherapy [\[16,17\].](#page-10-0) This approach has the capacity

to specifically kill cancer cells and can potentially lead to the complete elimination of residual cancer cells [\[16\]](#page-10-0).

In the early stages (late 1980s), adoptive cell transfer based approaches used peripheral blood mononuclear cells exposed to IL-2 ex vivo to lead to the formation of what are called lymphokineactivated killer (LAK) cells, to kill cancer cells [\[18,19\].](#page-10-0) Although some encouraging responses were originally obtained in human subjects, considerable toxicity was seen with LAK cells and IL-2 treatment $[20-24]$ $[20-24]$ $[20-24]$, and adoptive immunotherapy with LAK cells did not become a practical option for the treatment of gynecological cancers.

A more sophisticated approach of adoptive cell transfer consists of ex vivo-stimulated TILs or tumorassociated lymphocytes from ascites, with or without IL-2 [\[25,26\]](#page-11-0).

Optimization of these adoptive immunotherapies will include improving the cell source, the forms of stimulation, the methods for in vitro expansion, and the cytokines that are given during such treatment.

2 Cancer vaccines: Either tumor-specific antigens (supplied as peptides) or antigen-activated dendritic cells are used, in combination with immune-stimulatory adjuvants, to stimulate T-cell responses [\[27\].](#page-11-0)

The best-known preventive vaccine is the one against the human papilloma virus (HPV), which is highly effective to prevent cervical cancer, vulvar cancer, oro-pharyngeal cancers, and anal cancers [\[28\].](#page-11-0) Therapeutic vaccines, on the other hand, are being developed in clinical trials, but no major breakthroughs have occurred to date.

3 Immune checkpoint inhibitors: Immune checkpoint proteins are expressed on cytotoxic T cells upon their activation, with the intention to keep the immune response under control, in a sort of "negative feedback loop" to hinder overactivation of the immune response and lessen damage to normal cells. Cancer cells utilize these pathways to evade immune surveillance. To counteract this undesired immune inhibition induced by the checkpoint proteins, monoclonal antibodies against these checkpoint proteins have been developed, including against the cytotoxic T lymphocytesassociated antigen 4 (CTLA-4), the programmed cell death (PD-1) protein, and the PD-1 ligand proteins (PDL-1), and are currently being used successfully in clinical settings $[29-31]$ $[29-31]$ $[29-31]$.

Side effects

The most common adverse reactions among patients treated with immune checkpoint inhibitors included fatigue (43%), musculoskeletal pain (27%), diarrhea (23%), pain and abdominal pain (22% each), and decreased appetite (21%). Eight percent of patients discontinue one of these drugs called pembrolizumab due to adverse reactions. Serious adverse reactions occurred among 39% of patients. The most frequent serious adverse reactions reported included anemia (7%), fistula (4.1%), hemorrhage (4.1%), and infections (4.1%) [\[31,32\].](#page-11-0)

Cervical cancer [\(Table 1\)](#page-3-0)

Cervical cancer is unique among gynecologic cancers because of its established quasi universal cause, human papillomavirus (HPV), the exact mechanisms whereby HPV proteins interact and disrupt the molecular machinery of the cell, and the existence of a pre-cancerous stage detectable by the pap smear. Of the many types of HPVs, more than 30 infect the genital tract. Of these, 14 are high-risk HPV subtypes, and two of the high-risk subtypes, 16 and 18, are found in up to 62% of cervical carcinomas and 82% of the precancerous diseases [\[28\]](#page-11-0). HPV-infected cancerous cervical epithelial cells express two of the viral genes, E6 and E7, that, respectively, interact with and disrupt the function of the p53 and retinoblastoma tumor suppressor gene products. Factors other than infection with HPV, such as cellular immune function, play an important role in determining whether the infection of cervical epithelial cells regresses or progresses to cancer. This has led to the development of prophylactic and therapeutic vaccines to HPV as well as treatment approaches based on the enhancement of host immune function.

Immunotherpy	Mechanism	Overall response rate
TA-HPV vaccine [54]	Viral vectors expressing HPV-16 or -18 E6 or E7	$20 - 50%$
Pembrolizumab [32]	Immune checkpoint inhibitors	<20%
Nivolumab [63]	Immune checkpoint inhibitors	$20 - 50%$
HPV-Tumor-infiltrating T cells $[65]$	Adoptive cell transfer	$20 - 50%$

Table 1 Immunotherapies for cervical cancer.

Vaccines

The biologic principle of cancer vaccines is to stimulate an immune response specifically directed against malignant cells. This can be applied prophylactically and therapeutically.

Prophylactic Human Papillomavirus Vaccine

For prophylactic vaccination, the goal is to induce an immune response that will recognize, eradicate, and prevent (pre-)malignant progression.

HPV E6 and E7 are attractive antigens for use in therapeutic vaccines due to the involvement of these HPV-encoded proteins in cellular transformation, and therefore, they are consistently expressed in HPV-positive tumor cells $[33-35]$ $[33-35]$. The development of prophylactic vaccination to protect against HPV infection became within the realm of possibility through the development of protein mimics that simulate the exterior protein capsid of the virus, termed virus-like particles (VLPs) [\[36\]](#page-11-0). Currently, there are three clinically available vaccines:

- 1 Gardasil (Merck, NJ, USA), a quadrivalent prophylactic vaccine containing the VLPs of four HPV types, 6, 11, 16, and 18. The vaccine was approved by the U.S. Food and Drug Administration (FDA) in 2006 [\[37\].](#page-11-0)
- 2 Cervarix (GlaxoSmithKline, Middlesex, U.K.), a bivalent prophylactic vaccine containing the virus-like proteins of types 16 and 18, that was approved in 2009 [\[38\]](#page-11-0).
- 3 Gardasil 9 (Merck, NJ, USA), a nonavalent vaccine containing the virus-like proteins of nine HPV types, 6, 11, 16, 18, 31, 33, 45, 52, and 58, that was approved by the U.S. FDA in 2016 [\[39\].](#page-11-0)

Clinical trials showed that both the bivalent and quadrivalent vaccines are highly efficient in preventing dysplastic and cancerous lesions, especially in patients who were not previously exposed to HPV. The efficacy of Gardasil was 99% for preventing cervical intraepithelial neoplasia grades 2 and 3 caused by HPV 16 or 18 in females who were not previously infected with either HPV 16 or 18 before vaccination; however, efficacy was reduced to 44% in those who were infected prior to vaccination $[40-42]$ $[40-42]$.

The bivalent and quadrivalent HPV vaccines were approved for females aged $9-26$ for cervical cancer prevention with an additional approval of the quadrivalent HPV vaccine for genital warts.

In late 2016, several months after its approval, the nonavalent HPV vaccine became the only available HPV vaccine in the United States given its trial efficacy and broader coverage of oncogenic HPV serotypes. However, globally, vaccine availability is still restricted and varies based on location and vaccine cost, with most countries adopting either the bivalent or quadrivalent HPV vaccine [\[43\]](#page-11-0).

Therapeutic Human Papillomavirus Vaccine

Cancer vaccines are being developed therapeutically to serve as a "booster" for pre-existing antitumor immune responses or activating antitumor immunotherapies that have been actively administered to the patient. Various therapeutic vaccine strategies have been explored, including live vector, nucleic acid, protein, whole cell, and combinatorial vaccines. However, currently, there

are no HPV therapeutic vaccines approved for use in humans. Nevertheless, there have been numerous and extensive studies that have generated promising vaccine candidates tested in clinical trials $[44-46]$ $[44-46]$ $[44-46]$.

Live vector vaccines

Live vector vaccines are recombinant bacterial or viral vectors that can replicate inside the host cells, facilitating the spread of antigens. They can drive antigen presentation through both MHC class I and class II pathways, stimulating $CD8^+$ cytotoxic T-cells and $CD4^+$ helper T-cells, respectively, thus provoking immune-mediated antitumor activity, and in cervical cancer, HPV-specific proteins have been utilized to target HPV-infected cells [\[47\].](#page-11-0)

Bacterial vector: Listeria is a promising vector because of properties such as its ability to infect macrophages without being captured by phagocytosis, and its ability to direct antigen processing via MHC I and MHC II pathways; therefore, Listeria monocytogenes (Lm) is of particular interest for vaccine development [\[48,49\]](#page-11-0). Promising results started to be published from various phase I, II, and III clinical trials, using bacterial vector vaccines against advanced cervical malignancies, reporting reduction in tumor size, increase in E7 cell-mediated immunity but only modest improvement in overall survival $[50 - 53]$ $[50 - 53]$ $[50 - 53]$.

Viral vectors: In clinical trials, recombinant modified vaccinia virus Ankara (MVA) viral vectors expressing HPV-16 or -18 E6 or E7 (TA-HPV) showed HPV-specific cytotoxic T cell (CTL) responses in 28% of patients with advanced cervical cancer in a phase I/II study [\[54,55\],](#page-12-0) and at least, a 40% reduction in lesions in 83% of patients aged $42-54$ with high-grade vulvar or vaginal intraepithelial neoplasia in a phase II study [\[56\]](#page-12-0). Most recently, a vaccine based on HPV-16 E2 (MVA E2) was shown to have 90% efficacy in the treatment of HPV-induced anogenital intraepithelial lesions in a phase III study in 1356 patients [\[57\]](#page-12-0).

However, there remain challenges in the use of live vector vaccines due to potential dominance of the immune response to the bacterial/viral vector instead of the HPV antigen and the potential pathogenicity of the vector, particularly among oncologic patients and immune-compromised individuals [\[47,58\].](#page-11-0)

Subunit vaccines

Various subunit vaccines have been explored and found to have an effect on cancer cells. Subunit vaccines are antigens delivered in the form of peptides or whole proteins. They are regarded as safer than live vector vaccines as they are present in the host cells transiently, decreasing the chances of toxicity $[59-61]$ $[59-61]$.

Immune checkpoint inhibitors

In cervical cancer, patients with advanced or disseminated recurrent disease have a poor prognosis and few substantial treatment options are available which make alternative treatment options such as immunotherapy attractive.

Preliminary results from the cervical cancer cohorts of several studies were recently published and included patients with recurrent squamous cell carcinoma and adenocarcinoma of the cervix. In these studies, treatment with anti-PD-1 therapy such as pembrolizumab and nivolumab demonstrated promising antitumor activity in patients with pretreated cervical cancer, without regard to tumor PD-L1 or other tumor biomarker expression [\[32,62,63\].](#page-11-0) Recently, based on studies, the FDA approved pembrolizumab (Keytruda, Merck and Co. Inc.) for patients with recurrent or metastatic cervical cancer with disease progression on or after chemotherapy [\[64\].](#page-12-0)

Adoptive cell transfer (ACT)

The efficacy of TIL therapy was demonstrated in cervical cancer by Stevanovic et al. [\[65\].](#page-12-0) TILs were extracted from cervical cancer tissue, expanded ex vivo, and selected for their ability to recognize the 102 E. Matanes, W.H. Gotlieb / Best Practice & Research Clinical Obstetrics and Gynaecology 60 (2019) 97-110

HPV-associated proteins E6 and E7. This approach resulted in two complete and one partial response in a clinical trial that included nine patients. At the time of publication, the two complete responses were ongoing 22 and 15 months after treatment, respectively.

After examining the T-cell response in patients who underwent complete regression, Stevanovic et al. surprisingly found that the reactive T cells were not directed against virally associated epitopes, but rather against cancer germline antigens (KK-LC1) or neoantigens (mutated SETDP1, METTL7) not previously recognized by the immune system $[66]$. These observations demonstrate one of the main challenges in T-cell-based therapy, which is the identification of those tumor associated antigens (TAAs) on tumors from individual patients that induce the highest level of T-cells activation at the tumor site.

Adverse effects:

Two main toxicities are associated with adoptive cell transfer:

- 1 Activation of infused tumor specific T cells can cause cytokine release syndrome $[67]$, which can lead to severe neurologic toxicities, hypoxia, and hypotension that requires immediate treatment with steroids or the anti-IL-6 receptor antibody tocilizumab [\[68,69\].](#page-12-0)
- 2 On-target toxicity of normal tissues that express the target antigen and off-target toxicity to tissues that express an unrecognized cross-reactive antigen [\[70\].](#page-12-0)

Ovarian cancer (Table 2)

Although >80% of patients with ovarian cancer will initially have a response to frontline treatment (surgery and platinum-based chemotherapy), the majority ultimately recur and eventually develop chemotherapy-resistant disease. The results of recent clinical trials, including trials that incorporate bevacizumab and poly adenosine diphosphate-ribose polymerase (PARP) inhibitors into chemotherapy regimens, suggest that a plateau has been reached for conventional therapies as there is no definitive increase in overall survival. Consequently, new treatment strategies and paradigms are of critical need for these patients.

Vaccines

Therapeutic vaccines for ovarian cancer have been developed and studied in various clinical trials. Currently, ovarian tumor vaccine approaches include:

- 1 Vaccination with defined tumor-associated antigens or DNA vaccines that encode for tumorassociated antigens.
- 2 Vaccination with whole tumor cell preparations, with and without the co-administration of antigen-presenting cells.

Specific tumor associated antigens

Peptide vaccines are designed to stimulate antitumor immune responses against various specific target antigens that are expressed on ovarian cancer, such as NY-ESO-1, p53, WT-1, HER-2, and VEGF [\[71,72\]](#page-12-0).

NY-ESO-1 antigen:

One of the candidate antigens for vaccine development is NY-ESO-1, which is a highly immunogenic cancer-testis antigen. It is expressed in up to 40% of ovarian cancers [\[73,74\].](#page-12-0) Two recent clinical trials have shown the ability of an NY-ESO-1 peptide vaccine with Montanide ISA51 as adjuvant to induce both antigen-specific CD4⁺ and CD8⁺ T cell responses in patients with minimal residual ovarian cancer [\[74\]](#page-12-0). The second trial used HLA-A*0201-restricted NY-ESO-1b peptide vaccination with Montanide in patients with ovarian cancer in complete clinical remission after first-line treatment [\[75\].](#page-12-0) Three of four patients with NY-ESO-1-positive tumors and four of five patients with NY-ESO-1-negative tumors showed T-cell immunity. Three patients with NY-ESO-1-negative tumors remained in complete clinical remission. Approximately 50% of patients showed NY-ESO-1-specific immune responses, resulting in a mean disease-free interval of 19.9 months [\[76\].](#page-12-0)

While these results should be interpreted with caution, the hypothesis that NYESO-1 targeted therapy may be associated with clinical benefit has become inevitable and would need to be confirmed in randomized clinical trials.

p53 peptides:

Other vaccination strategies have used p53 peptides as the immunogenic antigen because the overwhelming majority of serous cancers overexpress p53. In general, p53 vaccination trials have resulted in the successful generation of p53-specific immune responses, and the vaccines have been well tolerated. Clinical responses have been demonstrated in up to 20% of patients, but these responses have not necessarily correlated with vaccine-mediated anti $p53$ immunity $[77-79]$ $[77-79]$.

Whole tumor antigen vaccines

Dissimilar to using specific tumor-associated antigen, whole tumor antigen vaccines can potentially provide an immune response to a wider range of tumor antigens. These vaccines can be created using autologous tumor lysates or tumor-derived RNA and are, in general, administered with adjuvants such as GM-CSF, Montanide ISA-51, or toll-like receptor agonists. Patients may have a better clinical response to whole tumor antigen vaccination [\[80,81\]](#page-13-0).

Dendritic cells

Dendritic cells are highly effective antigen-presenting cells and play a central role in the induction of both CD4 and CD8 T cell responses. Dendritic cells can be pulsed with tumor antigen peptides or can be produced to express tumor antigens, allowing their use to enhance antitumor immunity. Exposure of T cells to dendritic cells pulsed with ovarian cancer-derived antigenic preparations has resulted in the generation of cytolytic effector T cells that are capable of killing autologous tumor cells in vitro [\[82\]](#page-13-0).

Monoclonal antibodies

Monoclonal antibodies can potentially induce antitumor responses in various ways:

1 Complement system activation

2 Interaction with tumor cell surface signaling molecules and inducing antiproliferative effects.

3 Enhancing the activity of phagocytic cells.

Bevacizumab

The only monoclonal antibody-based drug, which is currently FDA-approved for the treatment of ovarian cancer, is bevacizumab (Avastin) [\[83\]](#page-13-0). Bevacizumab is a recombinant-humanized monoclonal antibody that inhibits vascular endothelial growth factor A (VEGF-A). The FDA approval is based on the phase III GOG-0218 trial, in which the bevacizumab regimen reduced the risk of disease progression or death by 38% compared with chemotherapy alone. The median progressionfree survival was 18.2 months versus 12.0 months, respectively (HR, 0.62; 95% CI, 0.52 -0.75 ; $P < .0001$ [\[84\]](#page-13-0).

Oregovomab

Oregovomab is a murine monoclonal antibody to CA125 that has been studied as a complementary treatment for ovarian cancer. In studies published in 2008–2009, Oregovomab showed survival benefit as maintenance therapy in patients with ovarian cancer after first-line treatment. A subgroup of patients with favorable prognostic factors had a significantly longer time to relapse compared to patients in the placebo group $[85-87]$ $[85-87]$ $[85-87]$. Oregovomab was also studied in combination with carboplatin and paclitaxel during the first-line treatment and might provide immune adjuvant properties in this setting [\[88\]](#page-13-0); however, these trials were published by one group a decade ago, and no further data have emerged.

Immune checkpoint inhibitors

Although immune checkpoint blocking antibodies have shown significant promise in mediating tumor regression in cervix, vulvar, and some endometrial cancers, the response rates in ovarian cancer have been modest. The first published data supporting checkpoint inhibitors as a potentially valuable therapeutic approach in ovarian cancer were observed in trials of the anti-PD-1 antibody nivolumab and the anti PD-L1 antibody BMS-93655 [\[31\]](#page-11-0). In the study reported by Hamanishi et al. [\[89\],](#page-13-0) a phase II trial of nivolumab (an anti-PD-1 antibody) in 20 patients with platinum-resistant epithelial ovarian cancer showed a response rate of 15% (3/20) and a disease control rate of 45% (9/20). In this study, two patients experienced a complete response. Although small, the presence of complete responses in platinum-resistant, heavily pretreated patients with an overall poor prognosis is promising.

Two additional immune checkpoint trials using avelumab (anti-PD-L1 antibody) and pembrolizumab (anti-PD-1 antibody) were presented at the 2015 ASCO annual meeting; Avelumab [\[90\]](#page-13-0) showed an objective response rate (ORR) of 9.7% and a stable disease (SD) rate of 44.4% in platinumresistant ovarian cancer patients [\[91\]](#page-13-0), and pembrolizumab was shown to have an 11.5% ORR and 34.6% SD rate in advanced ovarian cancer with positive PD-L1 status [\[92\].](#page-13-0)

The results of the KEYNOTE-028, a Phase-1b study of pembrolizumab in 26 heavily pre-treated ovarian cancer patients, showed one complete response, two partial responses, and six patients with SD, corresponding to a disease control rate of 34.6% [\[93\].](#page-13-0)

Ongoing or planned phase 3 trials in ovarian cancer with immune checkpoint inhibitors include NCT02718417 (Javelin Ovarian 100), ENGOT-ov29-GCIG (ATALANTE), NCT02580058 (Javelin Ovarian 200), and NRG-GY009. These trials are testing combinations with chemotherapy and/or bevacizumab, or the potential efficacy as maintenance therapy.

Immune checkpoint inhibitors are further being investigated among BRCA mutation carriers suffering from ovarian cancer, in combination with PARP inhibitors. Specifically, a phase I study of olaparib (PARP inhibitor) and durvalumab (anti-PD-L1) found an ORR of 17% and a disease control rate of 83% [\[94\]](#page-13-0).

Adoptive cell therapy (ACT)

As with other immunotherapy approaches, ACT has been most broadly utilized in melanoma where response rates around 50% have been reported [\[95\].](#page-13-0) Furthermore, CAR-T cells have also demonstrated encouraging results of inducing complete response in 70%–90% of patients with relapsed or refractory B-cell acute lymphoblastic leukemia [\[96\].](#page-13-0) When it comes to ovarian cancer, there are only few reports regarding the efficacy of ACT, and some of them are promising; when ovarian cancer TIL therapy was administered after surgery and primary adjuvant chemotherapy, the results showed 100% 3-year survival in patients who received TIL versus 67.5% in those who did not [\[97\]](#page-13-0).

There are several ongoing trials evaluating TILs in ovarian cancer (NCT02482090, NCT01883297). Phase I studies of engineered T cells targeting MUC16 (NCT02498912), mesothelin (NCT01583686), and NY-ESO-1 (NCT01567891, NCT02457650) are ongoing. Although remarkable responses have been observed, most clinical responses are short-lived with eventual tumor relapse.

In summary, the data suggest a strong biological and clinical rationale for testing TIL therapy in ovarian cancer, but further clinical trials are needed.

Endometrial cancer (Table 3)

Endometrial cancer ranks as the most common gynecological cancer [\[98\]](#page-13-0). Early diagnosis and the prognosis are often very good from this increasingly common malignancy, but unfortunately, minimal progress has been made to improve survival for women suffering from advanced or recurrent disease with poor outcome [\[99\]](#page-13-0).

Immune checkpoint inhibitors

Programmed cell death-1 (PD-1) and its ligand PD-L1 are expressed in up to 80% of primary endometrial cancers and are expressed in as much as 100% of metastatic cases [\[100\]](#page-13-0). Additionally, microsatellite instability-high (MSI-H) tumors have been found to correlate with higher expression of PD-1 and PD-L1 [\[101\].](#page-13-0)

At the 2017 annual meeting of the American Society of Clinical Oncology (ASCO), new data from the phase Ib KEYNOTE-028 trial were presented evaluating anti-PD-1 (pembrolizumab) therapy in PD-L1 positive solid tumors and included a subgroup of patients with advanced endometrial cancer (MSI testing was not performed) [\[102\]](#page-13-0). Patients in this study had failed standard therapy and had received at least two prior lines of treatment. The tumors were required to have positive PD-L1 expression defined as at least 1% positive staining by immunohistochemistry. Out of 24 patients, three (12.5%) had a partial response and three had SD. This group included patients with all histologies and only one patient had an MSI-H tumor. Overall, pembrolizumab was well tolerated and there was no discontinuation of therapy due to toxicity. Preliminary findings of a phase Ib/II study of pembrolizumab and lenvatinib, a VEGF receptor kinase inhibitor, in 23 patients with metastatic endometrial cancer demonstrated an impressive ORR of 48% [\[103\].](#page-13-0)

In 2017, subsequent to the publication of the study by Le et al. [\[104\],](#page-13-0) showing the high sensitivity of MMR-deficient cancers to immune checkpoint blockade with objective radiographic responses in 53% of patients and complete responses in 21% of patients, pembrolizumab was FDA-approved for use in MSI-H or MMR-deficient solid tumors that have progressed on standard therapy and have no alternate therapeutic option, becoming the first drug to gain FDA approval based on genomic characterization rather than tumor site.

Table 3

Immunotherapies for endometrial cancer.

As we await the final results from these trials, additional early phase studies are currently recruiting patients to further assess the clinical benefit in women with advanced and recurrent endometrial cancer:

- (NCT02549209): Pembrolizumab in combination with standard treatment (carboplatin and paclitaxel) for patients with recurrent or advanced disease.
- (NCT02899793): Pembrolizumab in patients with polymutated, hypermutated, or MSI-H tumors, as this group was shown to be a promising targeted cohort.
- (NCT02982486): Combination immune checkpoint inhibitors (anti-PD-1 (Nivolumab) and anti-CTLA-4 (Ipilimumab) are also being investigated in patients with advanced grade 3 endometrial cancers and high-risk histologies (serous, clear cell, and mixed histology).

Summary

Immunotherapies represent a novel approach to treat gynecological cancers and offer the potential for extended benefits even in advanced disease. Currently, antibodies against immune checkpoints are the most common representatives of this treatment modality. An improvement in the understanding of the immune system in tumor immunosurveillance has resulted in the development of a new generation of immunotherapeutic agents. Combination with chemotherapy, molecular-targeted therapy and radiotherapy could also be viable treatment options. Many clinical trials are ongoing and will eventually give further insight into immunotherapy's proper place in gynecological cancers treatment.

Practice points

Immunotherapy is utilized to suppress the well-documented inhibitory effect of the tumor on the immune system and restore it to a state of activity, utilizing:

Monoclonal antibodies: The only monoclonal antibody-based drug which is currently FDAapproved for the treatment of ovarian cancer is bevacizumab (Avastin).

Cancer vaccines: Tumor-specific antigens (supplied as peptides) or antigen-activated dendritic cells, in combination with immune-stimulatory adjuvants, to stimulate T cell responses against cancer. Anti-HPV bivalent and quadrivalent vaccines are highly efficient in preventing dysplastic and cancerous lesions, especially in patients who were not previously exposed to HPV.

In addition, bacterial vector vaccines have been developed against cervical malignancies, and reported reduction in tumor size, increase in E7 cell-mediated immunity, but only modest improvement in overall survival.

Checkpoint inhibitors: To counteract the immune inhibition induced by the checkpoint proteins, monoclonal antibodies against these checkpoint proteins have been developed, including against the cytotoxic T lymphocyte associated antigen 4 (CTLA-4), the programmed cell death (PD-1) protein, and the PD-1 ligand proteins (PDL-1), and are currently being used in clinical settings.

In May 2017, the checkpoint inhibitor Keytruda^R (pembrolizumab) was approved by the FDA for the treatment of patients with unresectable or metastatic solid tumors including endometrial, which have been identified as having a biomarker referred to as MSI-H or mismatch repair deficient (dMMR)." In June 2018, the FDA granted a full approval to KEYTRUDA^R for patients with recurrent or metastatic cervical cancer with disease progression on or after chemotherapy whose tumors express PD-L1. Response rates, including complete responses, have been described with other anti-PD-1 antibodies in patients with platinum-resistant epithelial ovarian cancer.

Adoptive cell transfer: Autologous T-cells activated and expanded ex-vivo from TILs or from peripheral blood from the patient, and infused back into the patient so these T cells will attack the cancer cells.

Research agenda

The thought of exploiting the immune system to treat cancer has been pursued for the last three decades. Only recently has this powerful strategy finally moved to the front stage of oncology. This huge advancement is the consequence of years of meticulos work by pioneering scientists.

Oncologists are rushing to amplify the use of immunotherapy to benefit more cancer patients. Hundreds of clinical trials are under way to see whether improved responses can be attained by combination therapy approaches. Understanding the dynamics of the response to immunotherapy will assist in finding ways to overcome immune suppression, and counter-regulation will lead to development of effective personalized targeted approaches to treat cancer as part of standard of care for cancer patients.

The aims of this chapter are to:

- ✓ To discuss the recent advancement in immunotherapies that include cancer vaccines, cell-based therapy, and immune checkpoint blockade.
- ✓ To summarize the current strategies used in clinical trials.
- ✓ To explore future directions for utilizing immune based therapies for long-lasting durable cure.

Conflict of interest statement

The authors declare no financial or personal relationships with other people or organizations that could inappropriately influence (bias) the content of this article.

References

- [1] [Ooi EH, Psaltis AJ, Witterick IJ, Wormald PJ. Innate immunity. Otolaryngol Clin North Am 2010;43:473](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref1)-[87 \[vii\]](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref1).
- [2] [Bonilla FA, Oettgen HC. Adaptive immunity. J Allergy Clin Immunol 2010;125:S33](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref2)-[40](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref2).
- [3] [Meek B, Speijer D, de Jong PT, de Smet MD, Peek R. The ocular humoral immune response in health and disease. Prog](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref3) [Retin Eye Res 2003;22:391](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref3)-[415](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref3).
- [4] [Mayer-Barber KD, Barber DL. Innate and adaptive cellular immune responses to Mycobacterium tuberculosis infection.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref4) [Cold Spring Harb Perspect Med 2015;5.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref4)
- [5] [Rottenberg M, Cardoni RL, Andersson R, Segura EL, Orn A. Role of T helper/inducer cells as well as natural killer cells in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref5) [resistance to Trypanosoma cruzi infection. Scand J Immunol 1988;28:573](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref5)-[82.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref5)
- [6] [Rich RR, elMasry MN, Fox EJ. Human suppressor T cells: induction, differentiation, and regulatory functions. Hum](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref6) Immunol 1986:17:369-[87.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref6)
- [7] [Wieczorek M, Abualrous ET, Sticht J, Alvaro-Benito M, Stolzenberg S, Noe F, et al. Major histocompatibility complex](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref7) [\(MHC\) class I and MHC class II proteins: conformational plasticity in antigen presentation. Front Immunol 2017;8:292](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref7). [8] [Ollila J, Vihinen M. B cells. Int J Biochem Cell Biol 2005;37:518](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref8)-[23](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref8).
- $[9]$ [Verschoor CP, Puchta A, Bowdish DM. The macrophage. Methods Mol Biol 2012;844:139](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref9)-[56.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref9)
- [10] [Souza-Fonseca-Guimaraes F, Adib-Conquy M, Cavaillon JM. Natural killer \(NK\) cells in antibacterial innate immunity:](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref10) angels or devils? Mol Med $2012:18:270-85$ $2012:18:270-85$.
- [11] Zhang JM, An J. Cytokines, inflammation, and pain. Int Anesthesiol Clin $2007;45:27-37$.
- [12] [Grulich AE, van Leeuwen MT, Falster MO, Vajdic CM. Incidence of cancers in people with HIV/AIDS compared with](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref12) [immunosuppressed transplant recipients: a meta-analysis. Lancet 2007;370:59](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref12)-[67.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref12)
- [13] [https://www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm581296.htm.](https://www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm581296.htm)
- [14] [Dunn GP, Old LJ, Schreiber RD. The three Es of cancer immunoediting. Annu Rev Immunol 2004;22:329](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref14)-[60.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref14)
- [15] [Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ. Natural innate and adaptive immunity to cancer. Annu Rev Immunol](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref15) [2011;29:235](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref15)-[71.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref15)
- [16] [Mantia-Smaldone GM, Corr B, Chu CS. Immunotherapy in ovarian cancer. Hum Vaccines Immunother 2012;8:1179](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref16)-[91.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref16)
- [17] [Motz GT, Coukos G. Deciphering and reversing tumor immune suppression. Immunity 2013;39:61](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref17)-[73](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref17).
- [18] [Steis RG, Urba WJ, VanderMolen LA, Bookman MA, Smith 2nd JW, Clark JW, et al. Intraperitoneal lymphokine-activated](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref18) [killer-cell and interleukin-2 therapy for malignancies limited to the peritoneal cavity. J Clin Oncol 1990;8:1618](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref18)-[29](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref18).
- [19] [Lenzi R, Edwards R, June C, Seiden MV, Garcia ME, Rosenblum M, et al. Phase II study of intraperitoneal recombinant](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref19) [interleukin-12 \(rhIL-12\) in patients with peritoneal carcinomatosis \(residual disease](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref19) < 1 cm) associated with ovarian [cancer or primary peritoneal carcinoma. J Transl Med 2007;5:66](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref19).
- 108 E. Matanes, W.H. Gotlieb / Best Practice & Research Clinical Obstetrics and Gynaecology 60 (2019) 97-110
	- [20] [Rosenberg SA, Lotze MT. Cancer immunotherapy using interleukin-2 and interleukin-2-activated lymphocytes. Annu](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref20) Rev Immunol 1986:4:681-[709](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref20).
	- [21] [Rosenberg SA, Lotze MT, Muul LM, Leitman S, Chang AE, Ettinghausen SE, et al. Observations on the systemic admin](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref21)[istration of autologous lymphokine-activated killer cells and recombinant interleukin-2 to patients with metastatic](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref21) [cancer. N Engl J Med 1985;313:1485](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref21)-[92.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref21)
	- [22] [Chiou SH, Sheu BC, Chang WC, Huang SC, Hong-Nerng H. Current concepts of tumor-in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref22)filtrating lymphocytes in human malignancies. J Reprod Immunol 2005:67:35-[50](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref22).
	- [23] [Berek JS, Lichtenstein AK, Knox RM, Jung TS, Rose TP, Cantrell JL, et al. Synergistic effects of combination sequential](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref23) [Immunoth](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref23) \tilde{A}^{\odot} [rapiesin a murine ovarian cancer model. Cancer Res 1985:4215](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref23)-[8.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref23)
	- [24] [West WH, Tauer KW, Yannelli JR, Marshall GD, Orr DW, Thurman GB, et al. Constant-infusion recombinant interleukin-2](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref24) [in adoptive immunotherapy of advanced cancer. N Engl J Med 1987;316:898](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref24)-[905.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref24)
	- [25] [Topalian SL, Solomon D, Avis FP, Chang AE, Freerksen DL, Linehan WM, et al. Immunotherapy of patients with advanced](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref25) cancer using tumor-infi[ltrating lymphocytes and recombinant interleukin-2: a pilot study. J Clin Oncol 1988;6:839](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref25)-[53.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref25)
	- [26] [Aoki Y, Takakuwa K, Kodama S, Tanaka K, Takahashi M, Tokunaga A, et al. Use of adoptive transfer of tumor-in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref26)filtrating [lymphocytes alone or in combination with cisplatin-containing chemotherapy in patients with epithelial ovarian cancer.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref26) Cancer Res 1[9](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref26)91:51:1934-9.
	- [27] [Lohmueller J, Finn OJ. Current modalities in cancer immunotherapy: immunomodulatory antibodies, CARs and vaccines.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref27) [Pharmacol Ther 2017;178:31](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref27)-[47.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref27)
	- [28] [Castellsague X, Diaz M, de Sanjose S, Munoz N, Herrero R, Franceschi S, et al. Worldwide human papillomavirus etiology](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref28) [of cervical adenocarcinoma and its cofactors: implications for screening and prevention. J Natl Cancer Inst 2006;98:](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref28) $303 - 15$ $303 - 15$ $303 - 15$
	- [29] [Phan GQ, Yang JC, Sherry RM, Hwu P, Topalian SL, Schwartzentruber DJ, et al. Cancer regression and autoimmunity](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref29) [induced by cytotoxic T lymphocyte-associated antigen 4 blockade in patients with metastatic melanoma. Proc Natl Acad](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref29) [Sci U S A 2003;100:8372](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref29)-[7.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref29)
	- [30] [Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, et al. Safety, activity, and immune correlates of](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref30) anti-PD-1 antibody in cancer. N Engl I Med $2012:366:2443-54$.
	- [31] [Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, et al. Safety and activity of anti-PD-L1 antibody in pa](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref31)tients with advanced cancer. N Engl J Med $2012;366:2455-65$ $2012;366:2455-65$.
	- [32] [Frenel JS, Le Tourneau C, O'Neil B, Ott PA, Piha-Paul SA, Gomez-Roca C, et al. Safety and ef](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref32)ficacy of pembrolizumab in [advanced, programmed death ligand 1-positive cervical cancer: results from the phase Ib KEYNOTE-028 trial. J Clin](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref32) [Oncol 2017;35:4035](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref32)-[41.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref32)
	- [33] [Brulet JM, Maudoux F, Thomas S, Thielemans K, Burny A, Leo O, et al. DNA vaccine encoding endosome-targeted human](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref33) [papillomavirus type 16 E7 protein generates CD4](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref33)+ [T cell-dependent protection. Eur J Immunol 2007;37:376](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref33)-[84.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref33)
	- [34] [Wu TC, Guarnieri FG, Staveley-O'Carroll KF, Viscidi RP, Levitsky HI, Hedrick L, et al. Engineering an intracellular pathway](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref34) [for major histocompatibility complex class II presentation of antigens. Proc Natl Acad Sci U S A 1995;92:11671](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref34)-[5.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref34)
	- [35] [Ji H, Wang TL, Chen CH, Pai SI, Hung CF, Lin KY, et al. Targeting human papillomavirus type 16 E7 to the endosomal/](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref35) [lysosomal compartment enhances the antitumor immunity of DNA vaccines against murine human papillomavirus type](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref35) 16 E7-expressing tumors. Hum Gene Ther $1999:10:2727-40$.
	- [36] [Chen XS, Garcea RL, Goldberg I, Casini G, Harrison SC. Structure of small virus-like particles assembled from the L1](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref36) protein of human papillomavirus 16. Mol Cell $2000:5:557-67$.
	- [37] [https://www.fda.gov/biologicsbloodvaccines/safetyavailability/vaccinesafety/ucm179549.htm.](https://www.fda.gov/biologicsbloodvaccines/safetyavailability/vaccinesafety/ucm179549.htm)
	- [38] [https://www.fda.gov/biologicsbloodvaccines/vaccines/approvedproducts/ucm186957.htm.](https://www.fda.gov/biologicsbloodvaccines/vaccines/approvedproducts/ucm186957.htm)
	- [39] <https://www.fda.gov/biologicsbloodvaccines/vaccines/approvedproducts/ucm426445.htm>.
	- [40] [Paavonen J, Naud P, Salmeron J, Wheeler CM, Chow SN, Apter D, et al. Ef](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref40)ficacy of human papillomavirus (HPV)-16/18 [AS04-adjuvanted vaccine against cervical infection and precancer caused by oncogenic HPV types \(PATRICIA\):](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref40) final analysis of a double-blind, randomised study in young women. Lancet $2009;374:301-14$ $2009;374:301-14$.
	- [41] [Quadrivalent vaccine against human papillomavirus to prevent high-grade cervical lesions. N Engl J Med 2007;356:](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref41) $1915 - 27.$ $1915 - 27.$ $1915 - 27.$
	- [42] [Romanowski B, Schwarz TF, Ferguson L, Peters K, Dionne M, Behre U, et al. Sustained immunogenicity of the HPV-16/18](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref42) [AS04-adjuvanted vaccine administered as a two-dose schedule in adolescent girls:](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref42) five-year clinical data and modeling [predictions from a randomized study. Hum Vaccines Immunother 2016;12:20](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref42)-[9](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref42).
	- [43] [St Laurent J, Luckett R, Feldman S. HPV vaccination and the effects on rates of HPV-related cancers. Curr Probl Cancer](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref43) [2018;42:493](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref43)-[506](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref43).
	- [44] [Vici P, Pizzuti L, Mariani L, Zampa G, Santini D, Di Lauro L, et al. Targeting immune response with therapeutic vaccines in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref44) [premalignant lesions and cervical cancer: hope or reality from clinical studies. Expert Rev Vaccines 2016;15:1327](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref44)-[36.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref44) [45] [Yang A, Farmer E, Wu TC, Hung CF. Perspectives for therapeutic HPV vaccine development. J Biomed Sci 2016;23:75.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref45)
- *[46] [Kim HJ. Current status and future prospects for human papillomavirus vaccines. Arch Pharm Res 2017;40:1050](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref46)–[63](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref46).
- [47] [Lin K, Doolan K, Hung CF, Wu TC. Perspectives for preventive and therapeutic HPV vaccines. J Formos Med Assoc 2010;](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref47) [109:4](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref47)-[24](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref47).
- [48] [Stevens R, Howard KE, Nordone S, Burkhard M, Dean GA. Oral immunization with recombinant listeria monocytogenes](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref48) [controls virus load after vaginal challenge with feline immunode](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref48)ficiency virus. I Virol 2004:78:8210–[8.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref48)
- [49] [Gunn GR, Zubair A, Peters C, Pan ZK, Wu TC, Paterson Y. Two Listeria monocytogenes vaccine vectors that express](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref49) [different molecular forms of human papilloma virus-16 \(HPV-16\) E7 induce qualitatively different T cell immunity that](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref49) [correlates with their ability to induce regression of established tumors immortalized by HPV-16. J Immunol 2001;167:](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref49) $6471 - 9$ $6471 - 9$ $6471 - 9$
- [50] Maciag PC, Radulovic S, Rothman J. The fi[rst clinical use of a live-attenuated Listeria monocytogenes vaccine: a Phase I](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref50) [safety study of Lm-LLO-E7 in patients with advanced carcinoma of the cervix. Vaccine 2009;27:3975](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref50)-[83](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref50).
- [51] [Skeate JG, Woodham AW, Einstein MH, Da Silva DM, Kast WM. Current therapeutic vaccination and immunotherapy](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref51) [strategies for HPV-related diseases. Hum Vaccines Immunother 2016;12:1418](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref51)-[29](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref51).
- [52] [Kawana K, Adachi K, Kojima S, Taguchi A, Tomio K, Yamashita A, et al. Oral vaccination against HPV E7 for treatment of](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref52) [cervical intraepithelial neoplasia grade 3 \(CIN3\) elicits E7-speci](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref52)fic mucosal immunity in the cervix of CIN3 patients. Vaccine $2014:32:6233 - 9$.
- [53] [Huh WBW, Dizon D, Powel MA, Leath CA, Landrum LM, Tanner E, et al. A prospective phase 2 trial \(GOG-0265\) of the](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref53) Listeria-based HPV immunotherapy axalimogene fi[lolisback \(AXAL\) in second- and third-line metastatic cervical cancer:](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref53) [an NRG Oncology Group trial. In: Society of gynecologic Oncology annual meeting. MD: National Harbor; 2017.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref53)
- [54] [Kaufmann AM, Stern PL, Rankin EM, Sommer H, Nuessler V, Schneider A, et al. Safety and immunogenicity of TA-HPV, a](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref54) recombinant vaccinia virus expressing modifi[ed human papillomavirus \(HPV\)-16 and HPV-18 E6 and E7 genes, in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref54) [women with progressive cervical cancer. Clin Cancer Res 2002;8:3676](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref54)-[85](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref54).
- [55] [Borysiewicz LK, Fiander A, Nimako M, Man S, Wilkinson GW, Westmoreland D, et al. A recombinant vaccinia virus](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref55) [encoding human papillomavirus types 16 and 18, E6 and E7 proteins as immunotherapy for cervical cancer. Lancet](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref55) [1996;347:1523](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref55)-[7.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref55)
- [56] [Baldwin PJ, van der Burg SH, Boswell CM, Offringa R, Hickling JK, Dobson J, et al. Vaccinia-expressed human papillo](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref56)[mavirus 16 and 18 e6 and e7 as a therapeutic vaccination for vulval and vaginal intraepithelial neoplasia. Clin Cancer Res](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref56) $2003:9:5205 - 13$ $2003:9:5205 - 13$
- [57] [Rosales R, Lopez-Contreras M, Rosales C, Magallanes-Molina JR, Gonzalez-Vergara R, Arroyo-Cazarez JM, et al. Regres](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref57)[sion of human papillomavirus intraepithelial lesions is induced by MVA E2 therapeutic vaccine. Hum Gene Ther 2014;](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref57) [25:1035](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref57)-[49](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref57).
- [58] [Medina E, Guzman CA. Use of live bacterial vaccine vectors for antigen delivery: potential and limitations. Vaccine 2001;](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref58) $19:1573 - 80$ $19:1573 - 80$
- [59] [Khong H, Overwijk WW. Adjuvants for peptide-based cancer vaccines. J Immunother Cancer 2016;4:56](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref59).
- [60] [Kenter GG, Welters MJ, Valentijn AR, Lowik MJ, Berends-van der Meer DM, Vloon AP, et al. Vaccination against HPV-16](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref60) [oncoproteins for vulvar intraepithelial neoplasia. N Engl J Med 2009;361:1838](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref60)-[47.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref60)
- [61] [Vallespi MG, Glaria LA, Reyes O, Garay HE, Ferrero J, Arana MJ. A Limulus antilipopolysaccharide factor-derived peptide](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref61) [exhibits a new immunological activity with potential applicability in infectious diseases. Clin Diagn Lab Immunol 2000;](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref61) [7:669](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref61)-[75.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref61)
- *[62] [Schellens JHM, Marabelle A, Zeigenfuss S, Ding J, Knowles Pruitt S, Chung HC. Pembrolizumab for previously treated](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref62) [advanced cervical squamous cell cancer: preliminary results from the phase 2 KEYNOTE-158 study. J Clin Oncol 2017;](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref62) [35\(suppl\). abstr 5514.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref62)
- [63] [Hollebecque A, Meyer T, Moore KN, Machiels JH, De Greve J, L](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref63)o[pez-Picazo JM, et al. An open-label, multicohort, phase I/II](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref63) [study of nivolumab in patients with virus-associated tumors \(CheckMate 358\): ef](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref63)ficacy and safety in recurrent or [metastatic \(R/M\) cervical, vaginal, and vulvar cancers. J Clin Oncol 2017;35\(suppl\). abstr 5504.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref63)
- [64] <https://www.fda.gov/drugs/informationondrugs/approveddrugs/ucm610572.htm>.
- [65] [Stevanovic S, Draper LM, Langhan MM, Campbell TE, Kwong ML, Wunderlich JR, et al. Complete regression of metastatic](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref65) [cervical cancer after treatment with human papillomavirus-targeted tumor-in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref65)filtrating T cells. J Clin Oncol 2015;33: $1543 - 50$ $1543 - 50$
- [66] [Stevanovic S, Pasetto A, Helman SR, Gartner JJ, Prickett TD, Howie B, et al. Landscape of immunogenic tumor antigens in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref66) [successful immunotherapy of virally induced epithelial cancer. Science 2017;356:200](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref66)-[5.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref66)
- [67] [Yu H, Sotillo E, Harrington C, Wertheim G, Paessler M, Maude SL, et al. Repeated loss of target surface antigen after](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref67) [immunotherapy in primary mediastinal large B cell lymphoma. Am J Hematol 2017;92:E11](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref67)-[3](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref67).
- [68] [Davila ML, Riviere I, Wang X, Bartido S, Park J, Curran K, et al. Ef](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref68)ficacy and toxicity management of 19-28z CAR T cell [therapy in B cell acute lymphoblastic leukemia. Sci Transl Med 2014;6:224ra225](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref68).
- [69] [Morgan RA, Yang JC, Kitano M, Dudley ME, Laurencot CM, Rosenberg SA. Case report of a serious adverse event following](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref69) [the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2. Mol Ther 2010;18:843](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref69)-[51.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref69)
- [70] [Wang X, Chang WC, Wong CW, Colcher D, Sherman M, Ostberg JR, et al. A transgene-encoded cell surface polypeptide](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref70) [for selection, in vivo tracking, and ablation of engineered cells. Blood 2011;118:1255](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref70)-[63.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref70)
- [71] [Disis ML, Schiffman K, Guthrie K, Salazar LG, Knutson KL, Goodell V, et al. Effect of dose on immune response in patients](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref71) [vaccinated with an her-2/neu intracellular domain protein](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref71)–based vaccine. J Clin Oncol 2004;22:1916–[25](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref71).
- [72] [Odunsi K, Jungbluth AA, Stockert E, Qian F, Gnjatic S, Tammela J, et al. NY-ESO-1 and LAGE-1 cancer-testis antigens are](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref72) [potential targets for immunotherapy in epithelial ovarian cancer. Cancer Res 2003;63:6076](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref72)-[83](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref72).
- [73] [Szender JB, Papanicolau-Sengos A, Eng KH, Miliotto AJ, Lugade AA, Gnjatic S, et al. NY-ESO-1 expression predicts an](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref73) [aggressive phenotype of ovarian cancer. Gynecol Oncol 2017;145:420](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref73)-[5.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref73)
- [74] [Odunsi K, Qian F, Matsuzaki J, Mhawech-Fauceglia P, Andrews C, Hoffman EW, et al. Vaccination with an NY-ESO-1](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref74) peptide of HLA class I/II specifi[cities induces integrated humoral and T cell responses in ovarian cancer. Proc Natl](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref74) [Acad Sci U S A 2007;104:12837](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref74)-[42](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref74).
- [75] [Diefenbach CS, Gnjatic S, Sabbatini P, Aghajanian C, Hensley ML, Spriggs DR, et al. Safety and immunogenicity study of](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref75) [NY-ESO-1b peptide and montanide ISA-51 vaccination of patients with epithelial ovarian cancer in high-risk](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref75) first [remission. Clin Cancer Res 2008;14:2740](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref75)-[8](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref75).
- [76] [Odunsi K, Matsuzaki J, Karbach J, Neumann A, Mhawech-Fauceglia P, Miller A, et al. Ef](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref76)ficacy of vaccination with re[combinant vaccinia and fowlpox vectors expressing NY-ESO-1 antigen in ovarian cancer and melanoma patients. Proc](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref76) Natl Acad Sci U S A 2012:109:5797-[802.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref76)
- [77] [Rahma OE, Ashtar E, Czystowska M, Szajnik ME, Wieckowski E, Bernstein S, et al. A gynecologic oncology group phase II](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref77) [trial of two p53 peptide vaccine approaches: subcutaneous injection and intravenous pulsed dendritic cells in high](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref77) [recurrence risk ovarian cancer patients. Cancer Immunol Immunother 2012;61:373](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref77)-[84.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref77)
- [78] [Leffers N, Lambeck AJ, Gooden MJ, Hoogeboom BN, Wolf R, Hamming IE, et al. Immunization with a P53 synthetic long](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref78) peptide vaccine induces P53-specifi[c immune responses in ovarian cancer patients, a phase II trial. Int J Cancer 2009;](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref78) $125:2104 - 13.$ $125:2104 - 13.$ $125:2104 - 13.$ $125:2104 - 13.$
- [79] [Leffers N, Vermeij R, Hoogeboom BN, Schulze UR, Wolf R, Hamming IE, et al. Long-term clinical and immunological](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref79) effects of $p53-SLP(R)$ vaccine in patients with ovarian cancer. Int J Cancer 20[12](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref79);130:105-12.
- 110 E. Matanes, W.H. Gotlieb / Best Practice & Research Clinical Obstetrics and Gynaecology 60 (2019) 97-110
	- [80] [Chiang CL, Kandalaft LE, Coukos G. Adjuvants for enhancing the immunogenicity of whole tumor cell vaccines. Int Rev](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref80) Immunol 2011:30:150-[82.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref80)
	- [81] [Buckanovich RJ, Facciabene A, Kim S, Benencia F, Sasaroli D, Balint K, et al. Endothelin B receptor mediates the endo](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref81)thelial barrier to T cell homing to tumors and disables immune therapy. Nat Med $2008:14:28-36$.
	- [82] [Santin AD, Hermonat PL, Ravaggi A, Bellone S, Pecorelli S, Cannon MJ, et al. In vitro induction of tumor-speci](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref82)fic human [lymphocyte antigen class I-restricted CD8 cytotoxic T lymphocytes by ovarian tumor antigen-pulsed autologous den](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref82)dritic cells from patients with advanced ovarian cancer. Am I Obstet Gynecol $2000:183:601-9$.
	- [83] [https://www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm610664.htm.](https://www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm610664.htm)
- *[84] [Burger RA, Brady MF, Bookman MA, Fleming GF, Monk BJ, Huang H, et al. Incorporation of bevacizumab in the primary](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref84) [treatment of ovarian cancer. N Engl J Med 2011;365:2473](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref84)-[83](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref84).
- *[85] [Berek JS, Taylor PT, Gordon A, Cunningham MJ, Finkler N, Orr Jr J, et al. Randomized, placebo-controlled study of](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref85) [oregovomab for consolidation of clinical remission in patients with advanced ovarian cancer. J Clin Oncol 2004;22:](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref85) $3507 - 16$ $3507 - 16$
- [86] [Berek JS, Taylor PT, Nicodemus CF. CA125 velocity at relapse is a highly signi](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref86)ficant predictor of survival post relapse: [results of a 5-year follow-up survey to a randomized placebo-controlled study of maintenance oregovomab immu](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref86)notherapy in advanced ovarian cancer. J Immunother $2008;31;207-14$ $2008;31;207-14$.
- [87] [Berek J, Taylor P, McGuire W, Smith LM, Schultes B, Nicodemus CF. Oregovomab maintenance monoimmunotherapy](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref87) does not improve outcomes in advanced ovarian cancer. J Clin Oncol $2009;27;418-25$ $2009;27;418-25$.
- [88] [Braly P, Nicodemus CF, Chu C, Collins Y, Edwards R, Gordon A, et al. The Immune adjuvant properties of front-line](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref88) [carboplatin-paclitaxel: a randomized phase 2 study of alternative schedules of intravenous oregovomab chemo](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref88)[immunotherapy in advanced ovarian cancer. J Immunother 2009;32:54](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref88)-[65](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref88).
- *[89] [Hamanishi J, Mandai M, Ikeda T, Minami M, Kawaguchi A, Murayama T, et al. Safety and antitumor activity of anti-PD-1](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref89) [antibody, nivolumab, in patients with platinum-resistant ovarian cancer. J Clin Oncol 2015;33:4015](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref89)-[22](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref89).
- [90] [Kelly K, Infante JR, Taylor MH, Patel MR, Wong DJ, Iannotti N, et al. Safety pro](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref90)file of avelumab in patients with advanced [solid tumors: a pooled analysis of data from the phase 1 JAVELIN solid tumor and phase 2 JAVELIN Merkel 200 clinical](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref90) trials. Cancer 2018:124:2010-[7.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref90)
- [91] [Disis ML, Patel MR, Pant S, Hamilton EP, Lockhart AC, Kelly K, et al. Avelumab \(MSB0010718C; anti-PD-L1\) in patients](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref91) [with recurrent/refractory ovarian cancer from the JAVELIN Solid Tumor phase 1b trial: safety and clinical activity. J Clin](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref91) [Oncol 2016;34.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref91)
- [92] [Varga A, Piha-Paul SA, Ott PA, Mehnert JM, Berton-Rigaud D, Johnson EA, et al. Antitumor activity and safety of pem](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref92)[brolizumab in patients \(pts\) with PD-L1 positive advanced ovarian cancer: interim results from a phase Ib study. J Clin](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref92) [Oncol 2015;33:5510.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref92)
- *[93] [Varga A, Piha-Paul S, Ott PA, Mehnert JM, Berton-Rigaud D, Morosky A, et al. Pembrolizumab in patients with pro](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref93)[grammed death ligand 1-positive advanced ovarian cancer: analysis of KEYNOTE-028. Gynecol Oncol 2018.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref93)
- [94] [Lee JM, Cimino-Mathews A, Peer CJ, Zimmer A, Lipkowitz S, Annunziata CM, et al. Safety and clinical activity of the](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref94) [programmed death-ligand 1 inhibitor durvalumab in combination with poly \(ADP-Ribose\) polymerase inhibitor ola](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref94)[parib or vascular endothelial growth factor receptor 1-3 inhibitor cediranib in women's cancers: a dose-escalation,](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref94) phase I study. J Clin Oncol $2017;35:2193-202$ $2017;35:2193-202$.
- [95] [Fujita K, Ikarashi H, Takakuwa K, Kodama S, Tokunaga A, Takahashi T, et al. Prolonged disease-free period in patients](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref95) [with advanced epithelial ovarian cancer after adoptive transfer of tumor-in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref95)filtrating lymphocytes. Clin Cancer Res 1995; $1:501 - 7.$ $1:501 - 7.$ $1:501 - 7.$
- [96] [Dudley ME, Wunderlich JR, Yang JC, Sherry RM, Topalian SL, Restifo NP, et al. Adoptive cell transfer therapy following](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref96) [non-myeloablative but lymphodepleting chemotherapy for the treatment of patients with refractory metastatic mel](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref96)[anoma. J Clin Oncol 2005;23:2346](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref96)-[57.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref96)
- [97] [Tomuleasa C, Fuji S, Berce C, Onaciu A, Chira S, Petrushev B, et al. Chimeric antigen receptor T-cells for the treatment of](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref97) [B-cell acute lymphoblastic leukemia. Front Immunol 2018;9:239](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref97).
- [98] [Lortet-Tieulent J, Ferlay J, Bray F, Jemal A. International patterns and trends in endometrial cancer incidence, 1978-2013.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref98) [J Natl Cancer Inst 2018;110:354](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref98)-[61.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref98)
- [99] [Dellinger TH, Monk BJ. Systemic therapy for recurrent endometrial cancer: a review of North American trials. Expert Rev](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref99) [Anticancer Ther 2009;9:905](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref99)-[16](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref99).
- [100] [Mo Z, Liu J, Zhang Q, Chen Z, Mei J, Liu L, et al. Expression of PD-1, PD-L1 and PD-L2 is associated with differentiation](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref100) [status and histological type of endometrial cancer. Oncol Lett 2016;12:944](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref100)-[50](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref100).
- *[101] [Makker V, Green AK, Wenham RM, Mutch D, Davidson B, Miller DS. New therapies for advanced, recurrent, and met](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref101)[astatic endometrial cancers. Gynecol Oncol Res Pract 2017;4:19](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref101).
- *[102] [Ott PA, Bang YJ, Berton-Rigaud D, Elez E, Pishvaian MJ, Rugo HS, et al. Safety and antitumor activity of pembrolizumab in](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref102) [advanced programmed death ligand 1-positive endometrial cancer: results from the KEYNOTE-028 study. J Clin Oncol](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref102) [2017;35:2535](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref102)-[41.](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref102)
- [103] [Makker V, Rasco DW, Dutcus CE, Stepan DE, Li D, Schmidt EV, et al. A phase Ib/II trial of lenvatinib \(LEN\) plus pem](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref103)[brolizumab \(Pembro\) in patients \(Pts\) with endometrial carcinoma. J Clin Oncol 2017;35](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref103).
- *[104] [Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, et al. Mismatch repair de](http://refhub.elsevier.com/S1521-6934(19)30032-X/sref104)ficiency predicts response of solid tumors to PD-1 blockade. Science $2017:357:409-13$.